

Thermal Decomposition of Mercerized Linter Cellulose and its Acetates Obtained from a Homogeneous Reaction

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Abstract: Cellulose acetates with different degrees of substitution (DS, from 0.6 to 1.9) were prepared from previously mercerized linter cellulose, in a homogeneous medium, using *N,N*-dimethylacetamide/lithium chloride as a solvent system. The influence of different degrees of substitution on the properties of cellulose acetates was investigated using thermogravimetric analyses (TGA). Quantitative methods were applied to the thermogravimetric curves in order to determine the apparent activation energy (E_a) related to the thermal decomposition of untreated and mercerized celluloses and cellulose acetates. E_a values were calculated using Broido's method and considering dynamic conditions. E_a values of 158 and 187 kJ mol⁻¹ were obtained for untreated and mercerized cellulose, respectively. A previous study showed that C6OH is the most reactive site for acetylation, probably due to the steric hindrance of C2 and C3. The C6OH takes part in the first step of cellulose decomposition, leading to the formation of levoglucosan and, when it is changed to C6OCOCH₃, the results indicate that the mechanism of thermal decomposition changes to one with a lower E_a . A linear correlation between E_a and the DS of the acetates prepared in the present work was identified.

Keywords: *Linter cellulose, cellulose acetates, thermal decomposition.*

Decomposição Térmica de Celulose de Linter Mercerizado e seus Acetatos Obtidos a partir de Reação Homogênea

Resumo: Acetatos de celulose com graus de substituição, GS, variando entre 0,6 e 1,9, foram preparados previamente a partir de celulose de linter mercerizado, em meio homogêneo, usando *N,N*-dimetilacetamida/cloreto de lítio como sistema de solvente. A influência de diferentes graus de substituição nas propriedades dos acetatos de celulose foi investigada usando a análise termogravimétrica (TGA). Métodos quantitativos foram aplicados nas curvas termogravimétricas obtidas a fim de determinar a energia de ativação aparente (E_a) relacionado à decomposição térmica de celulose não-tratada e mercerizada e acetatos de celulose. Valores de E_a foram calculados usando o método de Broido e considerando condições dinâmicas. Valores de E_a de 158 e 187 kJ mol⁻¹ foram obtidos para a celulose não-tratada e mercerizada, respectivamente. Em trabalho anterior verificou-se que o C6OH é o sítio mais reativo na acetilação, provavelmente devido ao impedimento estérico de C2 e C3. O C6OH participa da primeira etapa de decomposição da celulose, levando à formação de levoglucosana e, quando se tem a substituição para C6OCOCH₃, o resultado indica que o mecanismo de decomposição térmica muda para um com E_a menor. Uma correlação linear entre E_a e o GS dos acetatos preparados no presente trabalho foi identificada.

Palavras-chave: *Celulose de linter, acetatos de celulose, decomposição térmica.*

Introduction

Cellulose, the most abundant naturally occurring polysaccharide, is renewable, biodegradable, and can be derivatized to yield various useful products^[1-3]. Cellulose is a linear homopolymer consisting of β -(1-4)-linked anhydroglucopyranose units (AGU). The wood is the main source of cellulose and it is considered a slowly regenerated raw material, because of the time required for a tree may be cut to produce cellulose^[4]. In contrast, cellulose from other raw materials, such as cotton, has become attractive as a source for cellulose derivatives, because it is available in large quantities, and is routinely cultivated around the world.

Cellulose is not soluble in conventional solvents, mainly due to the complex morphology of the crystalline regions and to inter- and intramolecular hydrogen bonds. The substitution of hydroxyl groups presents of the cellulose chains by less polar groups, aiming at the solubilization after derivatization, is very usual^[5]. The aprotic polar solvent (*N,N*-dimethylacetamide) in combination with an inorganic salt, preferably lithium chloride (LiCl)^[6], is able to dissolve cellulose. The impetus behind investigating this solvent system for cellulose derivatization under homogeneous reaction has been extensively described in the literature^[7-13]. The homogeneous reaction involves several steps, including cellulose

activation, dissolution, and subsequent reaction with a derivatizing agent^[10].

The derivatization of cellulose in a homogeneous reaction, as considered in the present work, is advantageous compared to a heterogeneous reaction, due to the ability to control the degree of substitution (DS) and regularity of substitution, both along the polymer backbone and among the three OH groups of the anhydroglucose unit (AGU). This reaction can be produced by employing materials with well-defined characteristics and through a method with good reproducibility, which allows the production a material with better properties^[10,12,14-17]. The cellulose from cotton linters used in the present study has a high degree of crystalline order, in consequence of this the polymer chains are closely packed and can only be dissolved in DMAc/LiCl after a pre-treatment with alkaline solution (mercerization)^[10].

The pre-treatment with alkali solution irreversibly convert the cellulose I into the second most extensively studied allomorph, cellulose II. The studies involving ¹³C CPMAS solid state NMR showed the highly crystalline structure of cellulose I and present the coexistence of two crystal phases: cellulose I α and cellulose I β . The crystalline phases I α e I β can occur in different amounts depending on the origin of the cellulose^[18].

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The cellulose I chains has a higher mobility, when swelled by NaOH, allowing them to rotate about their axes, producing the antiparallel structure, characteristic of cellulose II^[19]. This change is due to the fact that the hydroxymethyl group (-CH₂OH) can assume different conformations, generating two different packing structures of cellulose chains in a microcrystal. The parallel chain structure, characteristic of cellulose I, occurs when the -adjacent chains have the CH₂OH groups in the same conformation. The antiparallel structure, characteristic of cellulose II, occurs when the adjacent chains have the -CH₂OH groups in different positions^[11,19].

The pre-treatment of cellulose with aqueous solutions of NaOH may form crystallites composed of parallel cellulose chains, incorporating the hydrate sodium and hydroxide ions and to form alkyl cellulose^[20]. The comprehension of structure of mercerized cellulose has been made using the different techniques, as synchrotron X ray analysis^[21] and X ray diffraction coupled with ¹³C CP-MAS NMR^[22]. These studies support the view that the cellulose I present the parallel mode and the cellulose II the antiparallel structure^[11].

Cellulose acetates are some of the most important cellulose esters due their multiple applications, such as in textiles (clothing and fabrics), high absorbency products (diapers, cigarette filters, and other filters), thermoplastics products (films and plastic instruments), food products (food packaging), cosmetics and pharmaceuticals (extended capsule/tablet release agents and encapsulating agents), and medical instruments (hypoallergenic surgical products)^[23,24].

The acetylation reaction of cellulose disrupts the intra- and intermolecular interactions within the cellulose chain. When the cellulose is derivatized in a homogenous reaction, the acetate groups in glucopyranose rings can be regularly introduced along the cellulose chains.

Thermogravimetric analysis (TGA) is widely used to investigate the thermal decomposition of polymers and determine some kinetic parameters, e.g., the rate of decomposition, *k*; the reaction order, *n*; and the *E_a*^[25-27]. The values of these parameters are of major importance in the identification of the mechanisms involved in polymer decomposition^[27,28] and as estimates to the thermal stability of polymers.

The comprehension of cellulose decomposition is important to biomass thermochemical conversion^[29]; however, the decomposition of cellulose and its derivatives is an extremely complex process, and generally consists of a series of reactions.

Due to the complexity of thermal decomposition of cellulose, studies have attempted to identify the kinetic parameters therein. The identification of the thermal decomposition temperature by TGA defines the upper limits of processing^[30] for materials. The usefulness of cellulose acetate can be strongly influenced by its degradation and stabilization^[31].

The present study investigates the preparation of cellulose acetates from cotton linters in a homogeneous medium (*N,N*-dimethylacetamide/lithium chloride), evaluates the thermal behavior of these cellulose acetates via TGA, and estimates their thermal decomposition apparent activation energy using Broido's equation under nitrogen and dynamic conditions.

Experimental

Materials and reagents

The source cellulose material used in this investigation was cotton linters (kindly provided by Industria Fibra S/A, Americana, São Paulo, Brazil) with low degree of polymerization (DP). *N,N*-dimethylacetamide (DMAc, Synth) was purified by distillation

from CaH₂ under nitrogen followed by storage over 4 Å molecular sieves. Acetic anhydride (Ac₂O, Synth) was also distilled from the appropriate drying agents P₂O₅ prior to use. Before the reaction, lithium chloride (LiCl, Synth) was placed in vacuum oven and dried for 3 hours at 200 °C, and stored in a desiccator.

Cellulose mercerization

The cellulose mercerization used in this study was established in a previous work^[11]. Mercerization of linters cellulose was achieved in a solution of 20% NaOH (1:50 w/w) for 1 hour at 0 °C^[32]. After this period, the alkali-swollen was washed with distilled water until it is no more trace of NaOH. The product was kept at room temperature for 24 hours and then dried under vacuum at 60 °C until constant weight was obtained.

Characterization of cellulose

Degree of polymerization (DP)

In order to measure the degree of polymerization (DP), the cellulose samples were solubilized in copper ethylene diamine solution (CUEN, 1:1 v/v), using an Ostwald viscometer. The average molar mass is given by $\overline{M}_v = \overline{DP} \times 162$, according to the TAPPI standard T230 om - 89^[33].

Crystallinity index

X ray diffraction analysis was used to evaluate the crystallinity index of untreated and mercerized celluloses. The diffractogram patterns were obtained in a VEB CARL ZEISS-JENA URD-6 Universal Diffractometer operating with CuKα ($\lambda = 1.5406 \text{ \AA}$) generated at 40 kV and 20 mA. The crystallinity index of samples was determined using the method based on Buschle-Diller and Zeronian equation, as described elsewhere^[32]:

$$I_c = 1 - \left(\frac{I_{\min}}{I_{\max}} \right) \quad (1)$$

where *I_{min}* is the minimum intensity of the 10 $\bar{1}$ lattice at 17° (amorphous region of native cellulose) or 101 lattice at 15° (non-crystalline region of mercerized cellulose), and *I_{max}* is the maximum intensity of the 002 lattice at 22° (attributed to the crystalline region of the sample).

Morphological surface analysis

Scanning Electron Microscopy (SEM) was carried out with a LEO 440 ZEISS/LEICA model, operating at accelerating voltage of 20 kV. The samples were covered with a thin layer of gold prior to analysis.

Thermogravimetric Analyses

TGAs were carried out in a Shimadzu TGA-50 instrument from samples consisting of untreated and mercerized linters cellulose. Approximately 5.0 mg of sample was weighed on a platinum pan and heated from room temperature to 600 °C at 20 °C/min under a nitrogen flow rate of 20 mL/min.

Dissolution of the mercerized linters cellulose

The dissolution procedure used in this study was established in a previous work^[10]. Standard round-bottomed flask of 250 mL of capacity was equipped with a stopcock, cylindrical funnel (without equilibration side arm), mechanical stirrer, and condenser closed with a stopper. Linters cellulose (2.0 g) and LiCl (5.0 g) were added into the flask. The flask was immersed in an oil bath, the pressure was reduced (2 mm Hg) and the oil bath temperature was heated from room temperature to 110 °C at 3 °C/min. The oil bath temperature was externally controlled (FE50RP controller, Flyever,

São Carlos, São Paulo, Brazil). The mixture was stirred under these conditions for 30 minutes. The vacuum pump was turned off and then slowly DMAc (100.0 mL) was added with vigorous stirring. After the addition of the DMAc under reduced pressure, the system was connected to dry nitrogen and the condenser was provided with a drying tube. The temperature was raised to 150 °C at 4 °C/min and the reaction mixture was stirred in dried nitrogen atmosphere for 90 minutes with mechanical stirrer and condenser with a drying tube. The system was then cooled to 36 °C at 1 °C/min.

Acetylation of the cellulose

The cellulose solution obtained after cooled was heated from room temperature to 110 °C at 3 °C/min. The system was maintained under reflux and a flow of dry nitrogen atmosphere. Acetic anhydride (Ac₂O) was added drop-wise, according to molar ratios to the anhy-droglucose units (AGU). The system was kept in this temperature (110 °C) for 4 hours, under stirring, reflux and nitrogen flow. After that, the system was cooled to room temperature and further the final product was purified by precipitation in methanol. The resulting precipitate was purified again by Soxhlet extraction in methanol solvent for 7 days. The cellulose acetates were dried at 50 °C under vacuum oven until constant weight.

Degree of substitution of cellulose acetates

Proton NMR spectra of cellulose acetates were recorded on a Bruker AC-200 spectrometer operating at 200 MHz at 80 °C using 392 scans. For this purpose, the cellulose acetates were dissolved in DMSO-d₆ at concentration of 10.0 mg mL⁻¹. By the addition of deuterated trifluoro-acetic into the sample solution it is possible shift the signal of residual water and hydroxyl protons to a lower field outside the spectral region of interest without affecting the chemical shifts of the glucose ring protons^[11,12,17,24,34].

Thermogravimetric analyses

The analyses were carried out as described for linters cellulose.

Thermal decomposition of linters cellulose and acetates

The kinetic of thermal decomposition is typically studied using dynamic and isothermal experiments. In dynamic methods, the samples are heated with a linear heating rate from room temperature until to reach the complete decomposition of material, while isothermal methods are realized in different intervals of time during with careful selection of different temperatures close to the decomposition temperature^[35]. In the case of untreated linters cellulose, the decomposition of cellulose can be initiated at temperatures lower than 370 °C, while the maximum temperature is around 400 °C.

In order to identify relevant kinetic parameters, particularly the activation energy, various methods can be applied. In kinetic analysis, the thermal decomposition of polymer can be described as occurring by a set of series of reaction and the degree of conversion is given as a function of time t for each isothermal experimental. It is defined by:

$$-\frac{d(1-\alpha)}{dt} = k(1-\alpha)^n \quad (2)$$

where n is the reaction order, which is adopted in the present study as first-order kinetics^[36] in dynamic conditions, and k is the rate constant, which is assumed to obey the Arrhenius laws (Equation 3) for the beginning of the reaction:

$$k = Ae^{-E_a/RT} \quad (3)$$

α is the degree of conversion:

$$\alpha = \frac{W_e}{W_o} \quad (4)$$

where W_e is the mass loss as volatile fragments eliminated from the polymer and W_o is the initial mass.

In the case of an isothermal experiment, MacCallum^[37] proposed the following expression for Equation 2:

$$-\frac{d(1-\alpha)}{dt} = kf(1-\alpha)^n \quad (5)$$

where f is a function that represents the reaction mechanism^[36].

The integration of Equation 5 and followed by substitution in Equation 2 results in:

$$f(1-\alpha) = Ae^{\frac{-E}{RT}} t \quad (6)$$

which rearranges to:

$$\frac{E_a}{RT} + \ln[f(1-\alpha)] - \ln A = \ln t \quad (7)$$

If several isothermal experiments are considered, a plot of degree of conversion vs. the reciprocal of the temperature of the experiment will result in a straight line the slope of which will be the E_a/R ^[35].

In a dynamic experiment, as in the present work, from the initial temperature (T_o), the temperature (T) changes linearly as a function of heating rate (u), during the time t, then:

$$T = T_o + ut \quad (8)$$

A combination of Equations 2, 3, and 8 gives:

$$\frac{d(1-\alpha)}{(1-\alpha)^n} = -\frac{A}{u} e^{\frac{-E}{RT}} dT \quad (9)$$

which, according to the Broido method^[38] can be written as:

$$\ln \left[\ln \left(\frac{1}{1-\alpha} \right) \right] = \frac{-E_a}{RT} + \ln \left[\left(\frac{R}{E_a} \right) \left(\frac{Z}{u} \right) T_m^2 \right] \quad (10)$$

where T_m is the temperature of the maximum reaction decomposition, Z is a constant, U is the heating rate, and R is the universal gas constant ($R = 8.314 \text{ J mol}^{-1} \text{ K}^{-1}$). Plotting $\ln[\ln[1/(1-\alpha)]]$ vs. $1/T$ should result in a straight line, the slope of which $-E_a/R$ can provide the value of E_a .

Results and discussion

Cellulose characterization

The scanning electron micrographs presented in Figure 1 show surfaces of mercerized and untreated cellulose linters.

The surface of mercerized linters is smoother than that of untreated linters, which indicates that there are no fragments stuck to the fibers, suggesting that some fragments have been eliminated during the mercerization procedure. In addition, the fibers bundles seem more separated in mercerized cellulose (Figure 1). Table 1 shows the degree of polymerization (DPv) and crystallinity index (Ic) of untreated and mercerized linters cellulose:

The alkali treatment resulted in a decrease in the degree of the polymerization (DPv) of the cellulose due to the hydrolysis of glycosidic bonds^[11]. Mercerization also results in higher liquid

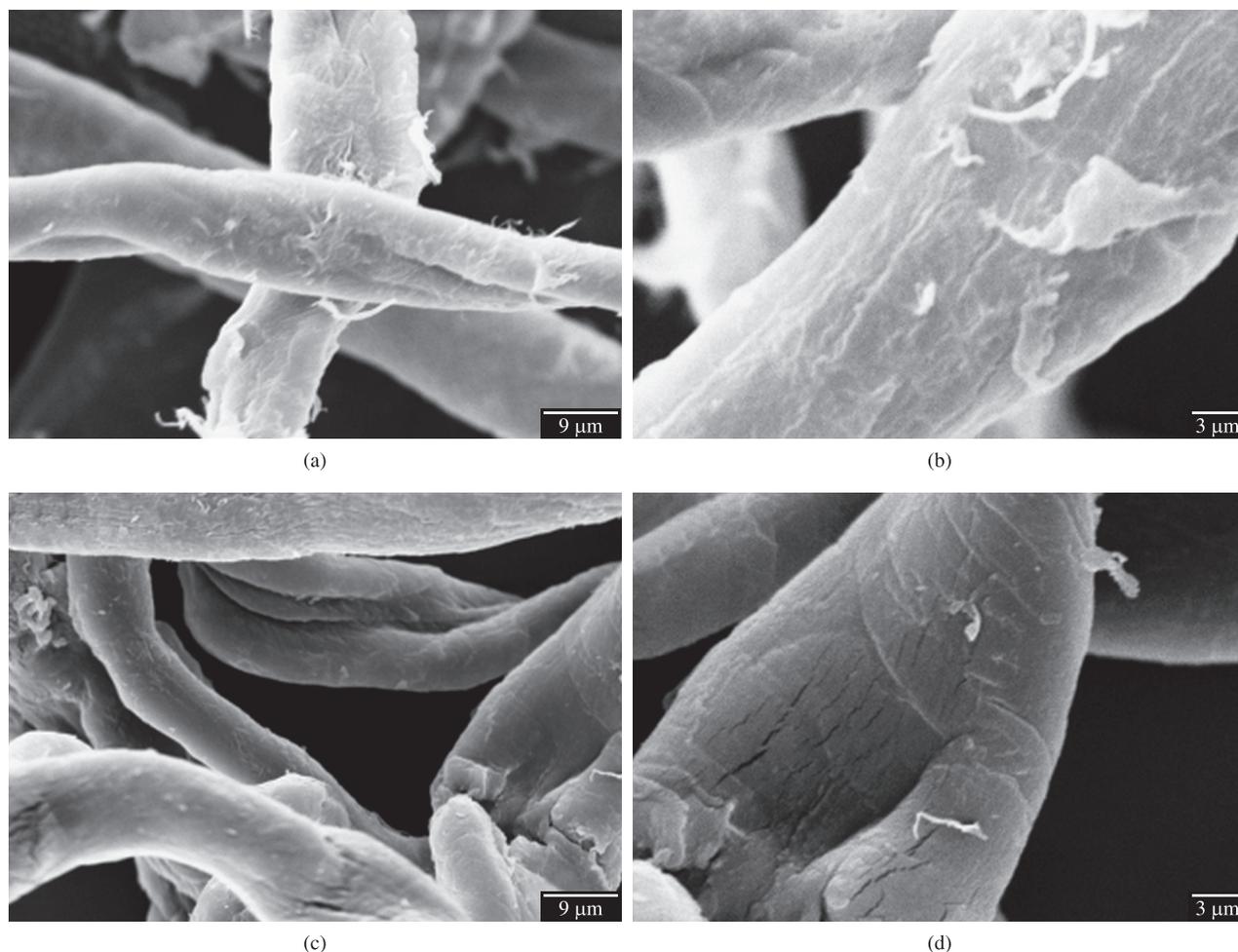


Figure 1. SEM images of untreated (a and b) and mercerized linters cellulose (c and d).

adsorption, as mercerization increases the specific surface area of the fiber, making the hydroxyl groups of the cellulose chains more accessible to solvents^[11,39]. In the present work, these changes enabled the dissolution of linters cellulose in DMAc/LiCl, the solvent system used for the acetylation reaction.

The penetration of NaOH seems to first start within the amorphous regions of cellulose, which are more accessible, and in a second step the non-crystalline region of cellulose, which leads to separation of part of the chains, decreasing the crystallinity. Treatment with 20% NaOH reduced the value of I_c from 79 to 73% (Table 1, X ray diffractogram not shown)

Characterization of cellulose acetates

The DS values of the cellulose acetates were calculated by means of the ratio between the area corresponding to the NMR proton resonance of the glucose ring (δ 2.60-5.20 ppm) and the corresponding resonance area of the methyl protons of the acetate group (δ 1.80-2.20 ppm), (spectra not shown)^[34]. The DS values of the cellulose acetates ranged from 0.6 to 1.9.

Even when a solution of cellulose appears visually homogeneous, there may be some degree of aggregation (irreversible process) and/or association (reversible process) of the dissolved polymer chains at a colloidal level. In this case, the cellulose chains are not in "true" solution, but homogeneous conditions of reactions are still preferable, as the distribution of the substituents along the chains will normally be more uniform than in a heterogeneous reaction. It is important to find the pretreatment and dissolution conditions

Table 1. Degree of polymerization (DP_v) and crystallinity index (I_c) of untreated and mercerized linters cellulose

Linters cellulose	DP _v	I_c (%)
Untreated	408	79
Mercerized	358	73

that maximize the molecular dispersion of the chains in solution, moreover because the phenomena of aggregation/association affect the efficiency of acetylation, since the hydroxyl groups that form H-bonds between chains will be much less available to react with Ac_2O ^[11]. The conditions used herein were based on previous studies^[10,11].

The X ray diffraction technique was employed to investigate the crystallinity of cellulose acetates (figures not shown). The results indicate that the I_c ranged from 33 to 65%, but there is no correlation between I_c and the degree of substitution of cellulose acetates; however, for all cellulose acetates, I_c decreased compared to mercerized linters cellulose ($I_c = 73\%$, Table 1). This reduction in crystallinity occurs due to the higher volume of the acetyl group introduced through acetylation, when compared to the hydrogen of hydroxyl groups, which hinders chain packing.

The thermal decomposition of cellulose and its acetates with different DS were investigated by TGA. Figure 2 shows DTG curves of the linters cellulose, mercerized linters cellulose, and cellulose

acetate with a DS of 0.8. The curves related to other acetates (figures not shown) present similar behavior.

The thermal decompositions of cellulose and its derivatives consist of a series of reactions. The first step of weight loss, from

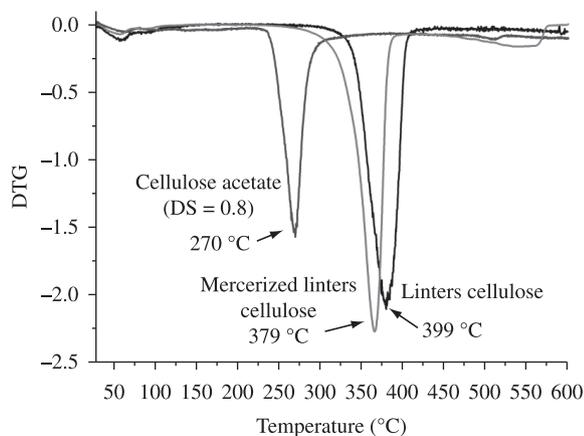


Figure 2. DTG curves of the linters cellulose, mercerized cellulose and cellulose acetate with a DS of 0.8 (atmosphere of N_2 20 mL/min, heating rate of 20 °C/min).

room temperature to 110 °C, is related to the evaporation of residual absorbed water. The second step ends near 450 °C, and can be described by an abrupt mass loss stage that is related to the main thermal decomposition process^[40,41].

The peak temperature in the DTG curves (Figure 2) corresponds to the temperatures at which the rate of decomposition reaches the maximum value^[41].

According to the results obtained, untreated cellulose begins to decompose at a higher temperature ($T_i = 368$ °C) when compared to mercerized cellulose ($T_i = 307$ °C). It is seen that there is a significant difference between mercerized and untreated cellulose in relation to their respective thermal decomposition temperatures, which may be related to the difference in crystallinity between untreated and mercerized linters. In the crystalline regions of the polymer, the intermolecular hydrogen bonds between chains are stronger than those of non-crystalline regions, and require more energy to break before the decomposition process can proceed. This means that the higher proportion of crystalline domains in the untreated cellulose results in a higher thermal stability, when compared to mercerized linters. The non-crystalline region is more actively decomposed than the crystalline region, adding complexities in the thermal decomposition of cellulose^[42].

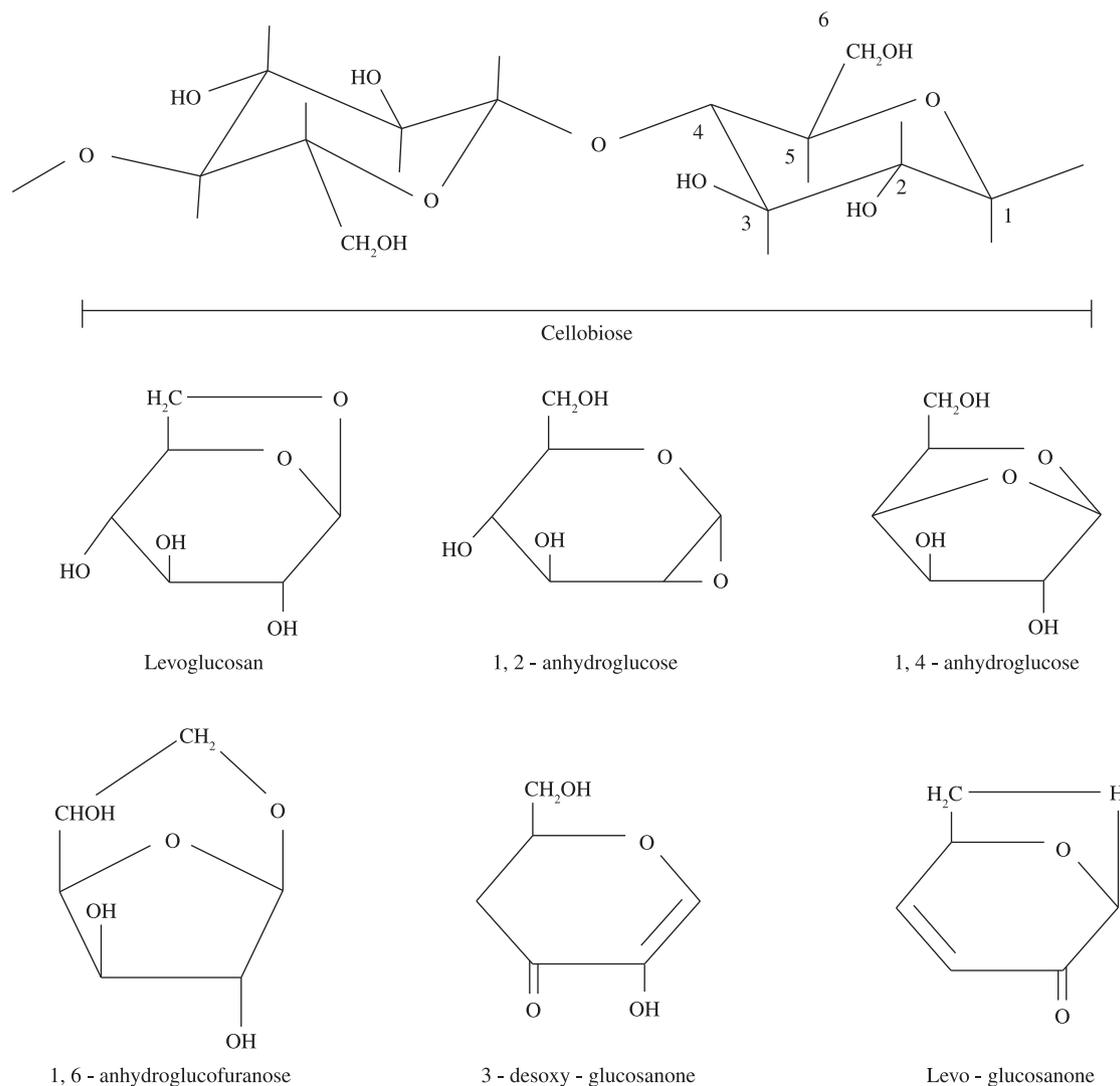


Figure 3. Anhydroglucoses and enones: Products of the thermal decomposition of cellulose^[41].

The first reaction in the decomposition of cellulose involves dehydration and leads to anhydrocellulose, which can result from either inter- or intra-ring dehydration^[43]. Figure 3 depicts the anhydroglucoses that can form as products of the thermal decomposition of cellulose.

The mechanism of the thermal decomposition of cellulose reveals the importance of the hydroxyl group connected to C6 in the decomposition process, which is involved in the reaction that leads the formation of levoglucosan (Figure 3).

Levoglucosan (1,6-anhydro- β -D-glucopyranose) is the primary product of cellulose pyrolysis; however, other anhydroglucoses (1,2-1,4-anhydro-glucose, 1,6-anhydroglucofuranose, and enones), furan, and furan derivatives are also produced^[44,45]. Levoglucosan formation from cellulose pyrolysis is produced by the scission of the 1,4 glucosidic linkage in the cellulose, followed by intramolecular rearrangement of the monomer units^[46].

Water formation from cellulose occurs at various temperatures because there is a number of different water producing reactions in the decomposition of this polysaccharide. Scheirs and co-workers^[43] showed some possibilities for the chemical elimination of water from cellulose that results in the formation of levoglucosan or 1,6-anhydroglucose. The thermostability of cellulose may be influenced by the introduction of acetates groups.

The plot of $\ln[\ln[1/(1-\alpha)]]$ vs. $1/T$ (Equation 10), in the temperature range that the main thermal decomposition step occurs (from 250 to 480 °C), is shown in Figure 4 for untreated and mercerized linters cellulose, as well as for cellulose acetate with a DS of 1.1. The E_a were determined from the slopes of the obtained linear curves (Figure 4)^[46]. Similar curves were obtained for the other cellulose acetates (figures not shown).

It can be seen that the E_a of untreated linters cellulose (158 kJ mol⁻¹) is lower than that of mercerized cellulose (187 kJ mol⁻¹). In cellulose I (untreated linters), whose chains are orientated in parallel, the -CH₂OH groups of adjacent chains have the same conformation, while in cellulose II (mercerized linters), the adjacent chains have these groups in different positions (antiparallel structure)^[11,18-21]. Despite the lower crystallinity of mercerized linters, due to the different -CH₂OH group orientations in cellulose II, the hydrogen bonds among their chains are stronger than those of the cellulose I chains (untreated linters), which results in cellulose II being a more stable structure compared to cellulose I. This configurational difference results in a higher E_a for the thermal decomposition of mercerized linters. Considering the E_a values obtained from the thermogravimetric analyses described in literature, it can be inferred that the data discussed herein are inside the range mentioned for celluloses obtained from different sources, that is, from 179.5 to 190.6 kJ mol⁻¹^[47] and from 140 to 155 kJ mol⁻¹^[48].

A correlation between the E_a of mercerized cellulose and its acetates is not so easy, because the presence of acetyl groups introduces the possibility of other reactions; however, taking into account that the C6 is the more reactive carbon under the conditions used in the present study^[11], and the importance of -C6OH in the thermal decomposition of the chain backbone, some considerations can be made.

The reactivity of OH in the C6 position is favored, when compared to C2 and C3, because this group is the least sterically hindered of the AGU unity^[49]. Acetates samples here described were prepared in DMAc/LiCl following previous study^[11], and the order of reactivity observed for all samples was C6>>C2>C3, confirming the higher reactivity of C6OH. The lower reactivity of C3OH, when compared to C2OH, can be related to the fact the first hydroxyl group can still be involved in the intramolecular hydrogen bond^[11].

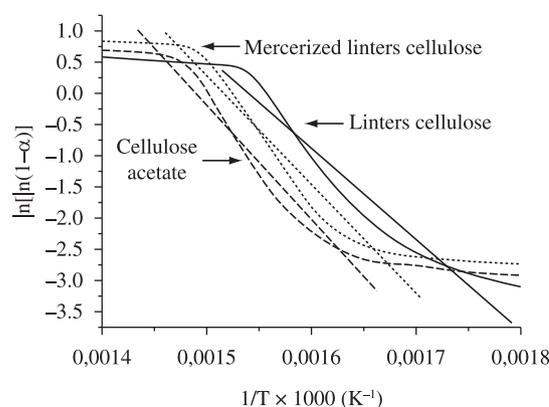


Figure 4. $\ln[\ln[1/(1-\alpha)]]$ vs. $1/T$ using Broido's equation for untreated and mercerized linters cellulose and cellulose acetate with a DS of 1.1.

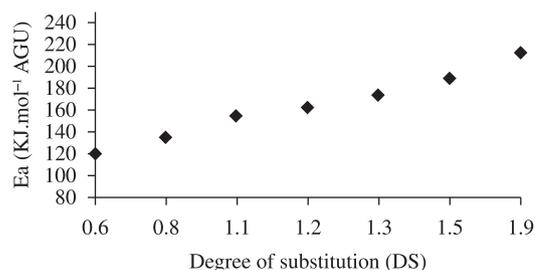


Figure 5. Apparent activation energy (E_a) vs. degree of substitution (DS) of cellulose acetates.

The C6OH takes part in the first step of cellulose decomposition, leading to levoglucosan formation (Figure 3). The E_a for mercerized cellulose (187 kJ mol⁻¹) decreased to the E_a for its acetates when the DS was varied from 0.6 to 1.2, wherein the E_a ranged from nearly 120 to 160 kJ mol⁻¹ (Figure 5). Considering that acetylation primarily occurs at C6^[11] from DS of 0.6 to 1.2, it can be considered that C6 is the most substituted when compared to C2 and C3^[11]. The obtained E_a suggest that thermal decomposition involving C6COOCH₃ can contribute to a lower E_a , when compared to levoglucosan formation from C6OH.

Considering the set of acetates, under the conditions considered in the present work there is a good correlation between E_a and DS, where E_a increases as a function of the DS (Figure 5). The range of investigated DS (from 0.6 to 1.9) was chosen to evaluate acetates with diversified substitutions on C6, C3, and C2.

As the DS increases, C2 and C3 increase their individual DS and their E_a increase. Even the acetate with a DS of 1.9 has a higher E_a (near 210 kJ mol⁻¹) than the mercerized cellulose. C2OH and C3OH take part in the thermal decomposition reactions that result, for instance, in 1,2-anhydroglucose and 3-desoxyglucosanone (Figure 3), respectively. When acetyl groups are introduced onto these carbons, the mechanism of thermal decomposition changes, and the values of E_a found in the present study indicate that the thermal decomposition involving C2COOCH₃ and C3COOCH₃ requires more energy, when compared to C6COOCH₃, contributing to a higher E_a .

Conclusions

Thermal decomposition behaviors and E_a values were evaluated by TGA and Broido's methods, respectively, for untreated and mercerized linters cellulose, as well as acetates synthesized in DMAc/LiCl from alkali treated samples (DS from 0.6 to 1.9), under dynamic conditions. The E_a values obtained for untreated

and mercerized celluloses, under the conditions used in the present study, indicate that the stronger hydrogen bonding present in the crystalline domains of cellulose II (mercerized linters) increase the corresponding Ea for thermal decomposition. The synthesis of acetates from low (0.6) to a higher (1.9) DS verified the influence of the distribution of the acetyl groups on C2, C3, and C6 on the Ea of thermal decomposition, under the conditions used in this study. It must be pointed out that the results herein provide important information regarding the thermal decomposition of untreated and mercerized linters cellulose, as well as that of their respective acetates; however celluloses and acetates from sources other than linters, or even linters with degrees of crystallinity different from the sample analyzed here, can present different behavior.

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References

- Wei, Y. & Cheng, F. - *J. Appl. Polym. Sci.*, **106**, p. 3624 (2007).
- Napolitano, B. A.; Martins, A. F.; Visconte, L. L. Y. & Nunes, R. C. R. - *Polímeros*, **14**, p.223 (2004).
- Corti, G. S.; Botaro, V. R.; Gil, L. F. & Gil, R. P. F. - *Polímeros*, **14**, p.313 (2004).
- Schurz, J. - *Prog. Polym. Sci.*, **24**, p.481 (1999).
- Botaro, V. R.; Santos, C. G. & Oliveira, V. A. - *Polímeros*, **19**, p.278 (2009).
- Ishii, D.; Tatsumi, D. & Matsumoto, T. - *Carbohydr. Res.*, **343**, p.919 (2008).
- Tosh, B.; Saikia, C. N. & Dass, N. N. - *Carbohydr. Res.*, **327**, p.5 (2000).
- Potthast, A.; Rosenau, T.; Sartori, J.; Sixta, H. & Kosma, P. - *Polymer*, **44**, p.7 (2003).
- Potthast, A.; Rosenau, T.; Buchner, R.; Roder, T.; Ebner, G.; Bruglachner, H.; Sixta, H. & Kosma, P. - *Cellulose*, **9**, p.41 (2002).
- Ramos, L. A.; Assaf, J. M.; El Seoud, O. A. & Frollini, E. - *Biomacromolecules*, **6**, p.2638 (2005).
- Ass, B. A. P.; Belgacem, M. N. & Frollini, E. - *Carbohydr. Polym.*, **63**, p.19 (2006).
- Ciaccio, G. T.; Ass, B. A. P.; Ramos, L. A. & Frollini, E. - *e-Polymers*, **22**, p.1 (2008).
- Liu, C. F.; Sun, R. C.; Zhang, A. P.; Ren, J. L.; Wang, X. A.; Qin, M. H.; Chao, Z. N. & Luo, W. - *Carbohydr. Res.*, **342**, p.919 (2007).
- Turbak, A. F.; El-Kafrawy, A.; Snyder, E. W. & Auerbach, A. B. U.S. - Patent 4.302.252 (1981).
- Turbak, A. F. - "Wood and agricultural residues", Soltes, E. J. (ed.), Academic Press, New York (1983).
- Klemm, D.; Philipp, B.; Heinze, T.; Heinze, U. & Wagenknecht, W. - "Comprehensive cellulose chemistry", in: *Functionalization of cellulose*, v.1, Wiley-VCH, Germany (1998).
- Ass, B. A. P.; Ciaccio, G. T. & Frollini, E. - *Bioresour. Technol.*, **97**, p.1696 (2006).
- Nishiyama, Y.; Sugiyama, J.; Chanzy, H. & Langan, P. - *J. Am. Chem. Soc.*, **125**, p.14300 (2003).
- Kroon-Batenburg, L. M. J. & Kroon, J. - *Glycoconjugate J.*, **14**, p.677 (1997).
- Shibasaki, H.; Kuga, S. & Okano, T. - *Cellulose*, **4**, p.75 (1997).
- Langan, P.; Nishiyama, Y. & Chanzy, H. - *Biomacromolecules*, **2**, p.410 (2001).
- Dinand, E.; Vignon, M.; Chanzy, H. & Heux, L. - *Cellulose*, **9**, p.7 (2002).
- Rinaudo, M. & Reguant, J. - "Natural polymers and agrofibers based composites", Frollini Leão, E. A. & Mattoso, L. H. C. (ed.), São Carlos, São Paulo, Brazil (2000).
- Paula, M. P.; Lacerda, T. M. & Frollini, E. - *Express Polym. Lett.*, **2**, p.423 (2008).
- Britto, D. & Campana-Filho, S. P. - *Thermochim. Acta*, **465**, p.73 (2007).
- Bozanic, D. K.; Djokovic, V.; Bibic, N.; Sreemukari Nair, P.; Georges, M. K. & Radhakrishnan, T. - *Carbohydr. Res.*, **344**, p.2383 (2009).
- Hanna, A. A.; Basta, A. H.; El-Saied, H. & Abadir, I. F. - *Die Angewandte Makromolekul. Chem.*, **260**, p.1 (1998).
- Levi, D. W.; Reich, L. & Tee, T. - *Polym. Eng. Sci.*, **5**, p.135 (1965).
- Shen, D. K. & Gu, S. - *Bioresour. Technol.*, **100**, p.6496 (2009).
- Sairam, M.; Sreedhar, B.; Mohan Rao, D. V. & Palaniappan, S. - *Polym. Advan. Technol.*, **14**, p.477 (2003).
- Lucena, M. C. C.; Alencar, A. E. V.; Mazzeto, S. E. & Soares, S. A. - *Polym. Degrad. Stab.*, **80**, p.149 (2003).
- Buschle-Diller, G. & Zeronian, S. H. - *J. Appl. Polym. Sci.*, **45**, p.967 (1992).
- "Viscosity of pulp (capillary viscosimeter method), T230 om-89", in: *TAPPI test methods*, Tappi Press, Atlanta (1990).
- Edgar, K. J.; Arnold, K. M.; Blount, W. W.; Lawniczak, J. E. & Lowmann, D. W. - *Macromolecules*, **28**, p.4122 (1995).
- Britto, D. & Campana-Filho, S. P. - *Polym. Degrad. Stab.*, **84**, p.353 (2004).
- Dahiya, J. B.; Kumar, K.; Muller-Hagedorn, M. & Bockhorn, H. - *Polym. Int.*, **57**, p.722 (2008).
- MacCallum, Jr. - "Thermogravimetric analysis", in: *Comprehensive polymer science*, Allen, G. & Bevington, L. C. (ed.), cap.37, Pergamon Press, Oxford (1989).
- Broido, A. - *J. Polym. Sci. Part A-2*, **7**, p.1761 (1969).
- Pusic, T.; Grancaric, A. M.; Sijacic, I. & Ribitsch, V. - *J. Soc. Dyers Colour.*, **115**, p.121, (1999).
- Huang, M-R. & Li, X-G. - *J. Appl. Polym. Sci.*, **68**, p.293 (1998).
- Flaqué, C. & Montserrat, S. - *J. Appl. Polym. Sci.*, **74**, p.201 (1999).
- Severian, D. - "Polysaccharides: structural diversity and functional versatility", Marcel Dekker, New York (2005).
- Scheirs, J.; Camino, G. & Tumiatti, W. - *Eur. Polym. J.*, **37**, p.933 (2001).
- Fengel, D. & Wegener, G. - "Wood: chemistry, ultrastructure, reactions", Walter de Gruyter, Berlin (1989).
- Li, S.; Lyons-Hart, J.; Banyasz, J. & Shafer, K. - *Fuel*, **80**, p.1809 (2001).
- Qu, X.; Wirsén, A. & Albertsson, A-C. - *Polymer*, **41**, p.4841 (2000).
- Kaloustian, J.; Pauli, A. M. & Pastor, J. - *J. Therm. Anal. Calorim.*, **63**, p.7 (2001).
- Milosavljević, I. & Suuberg, E. M. - *Ind. Eng. Chem. Res.*, **34**, p.1081 (1995).
- Marson, G. A. & El Seoud, O. A. - *J. Appl. Polym. Sci.*, **74**, p.1355 (1999).

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