

Nanofibers of gelatin and polivinyl-alcohol-chitosan for wound dressing application: fabrication and characterization

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Abstract

Electrospun nanofibers from gelatin (G), chitosan (CS), and chitosan-polyvinyl alcohol (CS-PVA) were developed by electrospinning process. Mechanical properties were determined by the tensile test, the elastic modulus values of the nanofibers were G (15.418-34.34 MPa) and CS-PVA (17.44-126.427 MPa). The morphological characterization by SEM revealed that the systems with 15% G and 6% CS-PVA showed morphological homogeneity. Structural characterization by FTIR indicated an interaction among some functional groups of the component. Thermal analysis by DSC and TGA showed degradation temperatures for G (330 °C), CS (210 °C to 370 °C), and PVA (310 °C to 420 °C). The contact angles values denoted the hydrophilic nature of the material. Finally, the antimicrobial assay proved that both 15% G and 7% PVA on the CS-PVA system presented the best antimicrobial effect. The results indicate that the electrospun nanofibers fabricated with G or CS-PVA can be used as wound healing dressings.

Keywords: *chitosan, electrospinning, gelatin, PVA, wound dressing.*

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1. Introduction

Nowadays, diabetes is the second cause of mortality in México, only 4.5% less than cardiovascular diseases. This metabolic disease affects glucose levels in blood, increasing them in consequence of a lack of insulin secretion or resistance^[1]. This prevalence is translated to a considerable economic impact in the country, where the National Health Sector has reported a cost of around 3,400 million dollars for the treatment of diabetes complications, above all, skin wounds^[2].

The reason which the health sector manages such a high budget resides on the complexity of the skin wound healing process in a diabetic patient. In comparison to a healthy person, the wound healing process of a diabetic patient is compromised^[3], consequence of multiple factors, such as immune system deficiency, poor circulation, metabolic disturbances, propensity of infection and loss of sensation because of neuropathy^[1]. On tissue injury, a healthy person forms a fibrin plug for either the re-establishment of homeostasis or the aggregation of platelets for the secretion

of growth factors (such as transforming growth factor-beta (TGF- β)^[4]). Subsequently, these inflammatory cells induce other growth factors, e.g. platelet-derived growth factor (PDGF) amongst others, all of this on the extracellular matrix (ECM)^[5]. Nevertheless, on a diabetic patient, the expression of these growth and angiogenic factors is impaired, stalling the healing process^[5,6].

Different alternatives for diabetic skin ulcer treatment, particularly on the polymeric material field, one of these alternatives is chitosan (CS), it is a natural polymer that has reported promising features for diabetic skin ulcers^[6,7], in fact there are some commercial wound dressings based on chitosan (Table 1). As addressed before, CS is a natural biopolymer, obtained by the partial deacetylation of chitin under strong alkaline conditions, formed by β (1-4)-D-glucosamine and β (1,4) N-acetyl-D-glucosamine (NAGA)^[8]. Aside from its antimicrobial, analgesic, antioxidant, and neuroprotective^[9-11] effects, chitosan has presented an effect upon the wound healing

Table 1. Commercial chitosan-based wound dressings.

Trademarks	Characteristics
Tegasorb P® 3M	Gel. Contains chitosan particles that swell while absorbing exudate and forms the gel.
Chitoflex® HemCom	Gel. Seals and stabilizes the wound
Chitopack C® Eisai	Cotton-like chitosan gel. Rebuilds body and subcutaneous tissue.

Modified from Liu et al.^[3]

process^[3], specifically on its effect of enhancing growth factor release^[10,12,13]. However, most of the reports of the chitosan effect upon wound healing of diabetic patients are based on gels, with the disadvantage that the treatment must be applied by a professional as well as quite often.

From different ways for chitosan processing, electrospinning for the obtaining of nanofibers is a way to surpass the previously mentioned gel disadvantages. These ultrafine continuous fibers are the product of high electric potentials^[14]. Apart from being a versatile (a great number of polymers can be processed), simple and the rather cheap process^[15], electrospun nanofibers possess a couple of properties which give them a great spectra of applications on biomedical materials: high surface area to volume ratio (which translates on oxygen permeability, fluid exchange without accumulation and uniform adherence *in situ*^[16]) accompanied with high porosity at the various pore size^[15]. There are reports of their application as scaffolds for different tissue regeneration^[17], cartilage^[18], bone^[19], drug delivery^[20] and, as the type of material of interest in the present work, wound dressings^[9]. This particular application is possible because of the nanofibers' structural capacity to attract the appropriate cellular growth substrates^[21]. As the principal barrier of protection for the wound, the scaffolds for wound dressing application have met some important criteria: They should facilitate gas permeation, present a controlled adhesion to the wound, as well as durability, flexibility and is capable of absorbing the wound exudates^[22]. Even though electrospun chitosan nanofibers possess many desirable characteristics, the electrospinning process for chitosan tends to be difficult, being that chitosan is polycationic and electrospinning principle is passed on charge application^[8]. Therefore, to process chitosan by electrospinning it requires the incorporation of other material to enable the process (electrospinning agent).

There are numerous electrospinning agents for chitosan, either natural or synthetic polymers. A natural polymeric alternative is a gelatin (G). This protein is prepared by collagen hydrolysis^[23], resulting in a structure consisting of mainly hydroxyproline, glycine, and proline amino acids^[24]. Aside from being biodegradable and electrospinning compatible, gelatin has the important characteristic of being affordable and giving electrospun fibers with controllable thickness and physical stability^[25]. On the other hand, some examples of synthetic biopolymers are polyethylene oxide (PEO)^[26], collagen^[27], and polycaprolactone (PCL)^[28]. However, all the previously mentioned electrospinning agents present disadvantages, such as the need for a crosslinking agent, difficult processing, and pearling formation. Another electrospinning agent for chitosan is polyvinyl alcohol (PVA) which is a

non-toxic and water-soluble synthetic biopolymer^[29], it has been reported a significant improvement on the mechanical properties of chitosan electrospun nanofibers^[30].

The aim of this study is to fabricate electrospun gelatin and chitosan-PVA nanofibers and evaluate their physicochemical properties (optical, mechanical, structural, thermal and morphological). An antimicrobial analysis was performed in order to explore the potential of the biomaterials produced for a possible application as wound dressings.

2. Materials and Methods

2.1 Materials

Gelatin (type B), medium weight chitosan with 75 to 85% deacetylated grade, and polyvinyl alcohol (PVA) (high Mw) were purchased from Sigma Aldrich, (St. Louis, Missouri, USA). Glacial acetic acid (GAA), CAS [64-19-7] with 2.5 pH and a density of 1.05 g/cm³ was obtained from Fagalah, MEX.

2.2 Preparation of G, CS, PVA and CS-PVA solutions

G solution was prepared following the method described by Okutan et al.^[31] with some modifications. The gelatin was dissolved in acetic acid solution (20% w/v) at 15 and 20% w/v. Solutions were stirred for 4 h, at 40 °C and 900 rpm on a magnetic stirrer until a clear and homogenous blend. CS solution was prepared at 1% w/v with the GAA and MiliQ water at 1:1 relation. The solution was stirred for 24 h at 25 °C. PVA solutions of 6%, 7%, and 8% w/v were prepared with MiliQ water as a solvent and were stirred at 900 rpm and 80 °C for 4 h. The CS-PVA solution was stirred at 900 rpm, 25 °C for 3 h at 1:1 relation.

2.3 Fabrication of G and CS-PVA nanofibers

The electrospinning process was carried at room temperature on a system composed as follows: A dual syringe infusion withdrawal pump (KDS 2010, KDScientific, Holliston, USA), a high voltage power supply (CZE1000R, Spellman, USA), the tip of the needle where the voltage was applied and an aluminum collector. Table 2 shows the sample number and the biopolymer used gelatin (G) or polyvinyl alcohol (PVA), as well as, the electrospinning conditions for G and CS-PVA. It is important to point out that in the case of CS-PVA solutions, CS % was a constant 1% w/v. PVA percentages were chosen based on their best morphological characteristics. Whereas CS percentage was the only one that could form nanofibers at the moment of electrospinning.

2.4 Characterization of G and CS-PVA nanofibers

2.4.1 Thickness

The thickness of the G and CS-PVA nanofibers were measured with a digital micrometer (Mitutoyo MDC -1"PX, Kawasaki-Shi, Kamagaya, Japan). To obtain the average thickness of the nanofibers, five measurements were performed, one measure on each corner and one in the middle of the resultant square nanofiber membranes.

Table 2. Formulations and electrospinning conditions of G and CS:PVA solutions.

Sample	G (%)	CS (%)	PVA (%)	Collector distance (cm)	Injection rate (mL/h)	Voltage (Kv)	Needle caliber (g)
1	15	-	-	10	1	-17	23
2	20	-	-	10	1	15	23
3	-	1	6	10	1	17	22
4	-	1	6	15	1	24	22
5	-	1	7	15	1	21	22
6	-	1	7	15	1	20	23
7	-	1	8	15	2	25	20
8	-	1	8	15	2	22	18

2.5 Color

The nanofibers color was evaluated with a colorimeter (Minolta CR-300, Osaka, Japan), calibrated with a standard ($Y= 94.1$, $x= 0.3155$, and $y= 0.3319$). Each nanofiber membrane was measured 5 times. The color change (ΔE), chromaticity (C^*), and Hue angle (H^*) were measured with Equations 1, 2, and 3, respectively.

$$\Delta E = \sqrt{(\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2} \quad (1)$$

$$C^* = \sqrt{a^{*2} + b^{*2}} \quad (2)$$

$$H^* = \text{artg} \left(\frac{b^*}{a^*} \right) \quad (3)$$

where L^* is luminosity; a^* is red/green coordinates; and b^* yellow/blue coordinates.

2.6 Tensile strength properties

The mechanical analysis of the nanofiber's membranes was measured using an universal tensile unit Shimadzu (AGS-X Kyoto, Japan) following the ASTM D1708. Rectangular membrane samples were cut with the following dimensions 5 mm wide and 24 mm long, the thickness of each membrane was measured in triplicate. Between 5 and 10 probes for each treatment were measured. The mechanical parameters, such as elastic modulus (EM), tensile strength (TS), and elongation at break (Eb) were calculated from the tensile test, the sample was clamped at the ends of the jaws of the equipment. The separation between clamps was 12 mm, 6 mm of headspace, at a head speed of 10 mm/min, and the load cell of 100 N was used for force measurements.

2.7 Morphological properties

Morphological properties of the electrospun nanofibers were evaluated by scanning electron microscopy (SEM), using a JEOL (JSM 3000, Akishima, Tokio) with a previous sample coating of Au/Pd using a Quorum Q150R. For better understanding, the sample number was delimited to 6 samples, 2 per PVA percentage, each one with different electrospinning conditions and one of 15% and 20% of gelatin each. A morphological analysis of the micromechanical tension

samples was focused on the fracture surfaces. In this case, an environmental scanning electron microscope (ESEM) FEI-Philips model XL30 ESEM (Tokio, Japan) was used, with a voltage of 20 kV and magnifications of 500X, 10000X, and 25000X.

2.8 Structural properties

Structural properties were characterized by Fourier Transform Infrared- Attenuated Total Reflectance (FTIR-ATR) spectroscopy (Thermo Scientific, with Nicolet iS5 with ATR, Waltham, USA) with a resolution of 4 cm^{-1} between 4000 and 400 cm^{-1} .

2.9 Thermal properties

The thermal properties were analyzed by differential scanning calorimeter (DSC) and thermogravimetric analysis (TGA). DSC analysis was performed using DSC from TA Instruments Inc. (Discovery Series, Delaware, USA) measurements were carried out under a nitrogen flow. Around 5 mg of sample was placed into aluminum cell and sealed, and an empty cell was used as reference. Once that the two cells were ready, these were placed inside the equipment, and heated at 10 $^{\circ}\text{C}/\text{min}$ from 25 $^{\circ}\text{C}$ to 300 $^{\circ}\text{C}$. TGA analysis was carried out on a thermogravimetric analyzer (TGA 8000 PerkinElmer Inc., Massachusetts, U.S.A.), the temperature range was from 24 $^{\circ}\text{C}$ to 700 $^{\circ}\text{C}$, at a heating rate of 10 $^{\circ}\text{C}/\text{min}$. This analysis was used to record the decomposition temperature of the nanofiber materials.

2.10 Surface hydrophilicity

The contact angle is a water contact angle of CS-PVA electrospun nanofibers was determined by Contact Angle Meter (CAM-Plus, ChemInstruments, Fairfield, USA). Ten measurements per PVA percentage and each membrane were taken. The droplet of water on a flat nanofiber (solid surface), the balance on the three-phase interface is expressed by Young's equation:

$$\gamma_s = \gamma_{sl} + \gamma_l \cos\theta \quad (4)$$

where the surface tension is γ_l , the contact angle between the interface liquid-air is θ , the interfacial tension γ_{sl} , and surface free energy of a solid is γ_s .

The sum of the interfacial tensions is given by γ_{sl} minus the work of adhesion, the work of adhesion can be expressed as:

$$W_A = \gamma_l (1 + \cos\theta) \quad (5)$$

2.11 Antimicrobial analysis

The nanofibers of G and CS-PVA were tested in their antimicrobial properties using a qualitative methodology reported by Ruiz-Ruiz et al.^[32] with some modifications. Müller-Hinton agar was inoculated with *S. tify* or *S. aureus* at 1×10^8 UFC and incubated at 37 °C for 12 h. Samples of 1.5 cm² were deposited on the surface of the culture and incubated at 37 °C for 8 h. A paper disk with ciprofloxacin was considered as control. Photographs of the resultant Petri dishes were taken by the imaging system Gel Doc™ (EZ system, BIO-RAD, Hercules, USA).

2.12 Statistical analysis

Statistical analyses of mechanical, optical, and contact angle characterization were processed by NCSS ver. 7 software (Kaysville, U.S.A). Data were presented by its mean ± standard deviation, at a significance level of P<0.05. If a significant difference was observed a mean comparison by Tukey-Kramer was processed.

3. Results and Discussions

3.1 Color

The color parameters ΔE, C*, and H* of the samples are presented in Table 3. In the case of ΔE, the behavior of the samples showed changes between those of G and CS-PVA. Samples 1 and 2 (G) showed higher ΔE values in comparison to samples 3 to 8 (CS-PVA). This behavior can be explained

Table 3. Optical properties of color of G and CS-PVA nanofibers.

Sample	ΔE	C*	H*
1	3.40 ± 0.17 ^c	1.33 ± 0.20 ^c	66.23 ± 2.01 ^d
2	2.02 ± 0.20 ^b	1.03 ± 0.08 ^c	82.38 ± 1.87 ^d
3	1.19 ± 0.41 ^{ab}	1.36 ± 0.39 ^{ab}	53.56 ± 3.77 ^b
4	1.42 ± 0.05 ^{ab}	1.33 ± 0.22 ^{ab}	49.14 ± 6.33 ^{bc}
5	0.68 ± 0.17 ^a	1.68 ± 0.74 ^a	61.16 ± 1.24 ^a
6	1.75 ± 0.10 ^b	1.06 ± 0.18 ^{bc}	61.33 ± 0.99 ^c
7	1.28 ± 0.44 ^{ab}	2.03 ± 0.31 ^{ab}	78.70 ± 1.45 ^{bc}
8	1.88 ± 0.12 ^b	1.92 ± 0.32 ^{cd}	76.24 ± 0.82 ^c

Data is presented as mean ± standard deviation. Abbreviation signification is follows: ΔE color difference, C* chromaticity, and H* Hue angle.

Table 4. Thickness and tensile at strength properties of G and CS-PVA nanofibers.

Sample	Thickness (mm)	EM (MPa)	TS (MPa)	E (%)
1	0.219 ± 0.05 ^b	34.342 ± 5.3 ^{b,c,d}	1.052 ± 0.318 ^{b,d}	4.178 ± 1.354 ^{a,c}
2	0.297 ± 0.073 ^b	15.418 ± 2.205 ^{b,d}	0.409 ± 0.215 ^{b,d}	4.344 ± 1.679 ^{a,c}
3	0.057 ± 0.014 ^a	NA ^a	14.89 ± 1.726 ^a	16.202 ± 1.56 ^a
4	0.032 ± 0.042 ^a	NA ^b	4.378 ± 3.063 ^{b,d}	16.852 ± 8.266 ^a
5	0.023 ± 0.003 ^a	116.49 ± 12.599 ^{b,c}	8.269 ± 2.252 ^{ab,c}	15.818 ± 5.693 ^a
6	0.035 ± 0.007 ^a	17.444 ± 2.948 ^{b,c,d}	4.237 ± 2.581 ^{b,d}	27.674 ± 9.698 ^{a,b}
7	0.024 ± 0.0005 ^a	29.201 ± 6.225 ^{b,c,d}	5.984 ± 3.426 ^{b,d}	23.155 ± 8.281 ^{a,b}
8	0.015 ± 0 ^a	126.427 ± 20.40 ^{b,c,d}	14.310 ± 5.047 ^{a,c}	26.502 ± 9.442 ^{a,b}

Data is presented as mean ± standard deviation. Different letters on each column indicate significant differences (P<0.05). Abbreviation signification is follows: EM: Elastic modulus; TS: Tensile strength; E: Elongation at brake.

with the reported by Horsfall^[33], the author concluded that the color change observed in the material is produced by the chemical composition of itself. The ΔE values for samples with gelatin revealed that the chemical composition would be strongly depended with the gelatin content in the samples, therefore a significant difference was found between them (P<0.05). However, this behavior becomes to be more complex in the case of the CS-PVA samples. Samples 5 and 6 are significantly different (P<0.05), even though they belong to the same PVA percentage. However, they differ in morphology, meanwhile sample 5 was fabricated with a needle caliber of 22 g, which has a higher diameter (0.7 mm) than the needle used on sample 6 which is caliber of 23 g and 0.6 mm of diameter. This effect can also be seen between samples 7 and 8.

Chromaticity can be defined as the color saturation. When a sample presents a low C* value, it infers that they present a high interference of colors, whereas a C* value of 0 means that the samples show an achromatic stimulus^[34]. The achromatic stimuli addressed previously represented itself by the visually white color of all the samples obtained. In contrast to ΔE values, samples 1 and 2 did not show chromatic differences between them, although these showed significant difference (P<0.05) with samples formulated with CS-PVA.

From previous optical parameters, H* is the most outwardly explaining. This parameter indicates the color of the sample within the color sphere, where tones are represented with the grades within the sphere. Being 90° represents yellow, 180° green, 270° blue and 360°, as well as 0°, represent red^[35]. According to Table 3, the samples were between 49.14° and 82.38°. This, along with a luminosity of average of 95 (data not shown) and the C* obtained, the samples presented a light color, in the yellow region, therefore, the whiteness present visually on all the samples.

3.2 Thickness

The thickness of G and CS-PVA nanofibers membranes are presented in Table 4, the values of these systems were found in the range from 0.219 to 0.297 mm and from 0.015 to 0.57 mm, respectively. The nanofibers of CS-PVA (samples 3 to 8) were obtained at different electrospinning conditions (Table 2), nevertheless, the thickness of these samples did not show significant differences (P>0.05) as it can be observed in Table 4. Although the electrospinning conditions experimented in this work were different, these allowed the fabrication of nanofibers at different percentages

of PVA. However, a significant difference was found between G and CS-PVA nanofibers, where G presented the highest values, a consequence of the different electrospinning conditions, added to the nature of the material, in this case, G can produce nanofiber with higher thickness. It is attributed that gelatin has the most complex and larger structure (formed by proteins, mineral salts, and water), samples 1 and 2 showed the higher nanofiber diameter with 111.66 and 286.33 nm (Table 5), respectively. It is difficult to compare our values with the literature because the references did not report the values and conditions that the nanofibers are produced.

3.3 Mechanical properties

The values of mechanical parameters (elastic modulus, tensile strength, and elongation at brake) for all the samples tested are presented in Table 4. As we can be observed, the samples 3 and 4 have not reported the EM data, as well as they present the higher standard deviation among all the samples. It is related to physical aspect of the material instead of the chemical nature. G nanofibers showed the lowest values in tensile strength and elongation at break, although these samples did not showed significant difference ($p > 0.05$) between them. The tensile strength values of samples of CS-PVA nanofibers is very fluctuating, it is attributed to

sample's thickness which is highly irregular (based on standard deviation). The elongation is a mechanical parameter that it is not related to the thickness. The elongation at break of CS-PVA samples increases as the PVA content is increased. Due to the thickness of the materials, it is difficult to compare these results with literature, since there is no information available (as far as the present authors know).

3.4 SEM analysis

SEM micrographs at 50000X of magnification of the electrospun CS:PVA nanofibers are presented on Figure 1, with their fiber diameter and general morphological characteristics presented on Table 5.

The samples of G had the highest fiber diameter value of 286.33 nm for 20% G, followed by 15% G with 111.66 nm. These results seem coherent since, as explained before, gelatin is a larger molecule. Also, G nanofibers of both concentrations (15% and 20% w/v) present the best morphological characteristics of all the materials since they do not present any drops or fractures. The last characteristics can be also observed in Figure 1a and 1b. In comparison with other studies^[36], the morphology of G nanofibers in the present study is also more homogeneous than gelatin nanofibers obtained in other studies^[31]. Following this comparison, it can be observed that gelatin concentration and voltage used in the electrospinning process were two times higher in the previous study than the used in the present study.

Samples 3 and 4 (6% PVA) showed small morphological improvement since fibrillar formation was observed, although they still presented fracture, peeling and drops. However, samples 5 and 6 presented the best morphological characteristics, since apart from fibrillar formation, they presented sparse fiber fracture, especially sample 6, which in comparison with sample 5, did not present pearling. Nevertheless, Samples 7 and 8 presented undesirable morphological characteristics again, with drops and fiber fracture, as well as an increase of the fiber diameter. The proportional increase in the nanofibers diameter has been reported by Tarus et al.^[37]. These authors reported the same effect on cellulose acetate nanofibers with a solvent system of acetone/dimethyl acetamide, where the diameter

Table 5. Nanofiber diameter and general morphological characteristics of G and CS-PVA nanofibers.

Sample	Fiber diameter (nm)	General morphological characteristics
1	111.66 ± 7.57 ^a	Fibrillar formation, no drops or fractures.
2	286.33 ± 2.3 ^b	Fibrillar formation, no drops or fractures.
3	33 ± 2.5 ^c	Fibrillar formation, drops and fiber fracture
4	33 ± 2.5	Fibrillar formation with fractures and pearling
5	17.8 ± 0.9 ^d	Fibrillar formation, pearling, sparse fiber fracture
6	17.8 ± 0.9	Fibrillar formation and sparse fiber fracture
7	27.8 ± 1.06 ^e	Fibrillar formation, drops and fiber fracture
8	27.8 ± 1.06 ^e	Fibrillar formation, drops and fiber fracture

Data are mean ± standard deviation ($n=10$). Significant difference ($P < 0.05$).

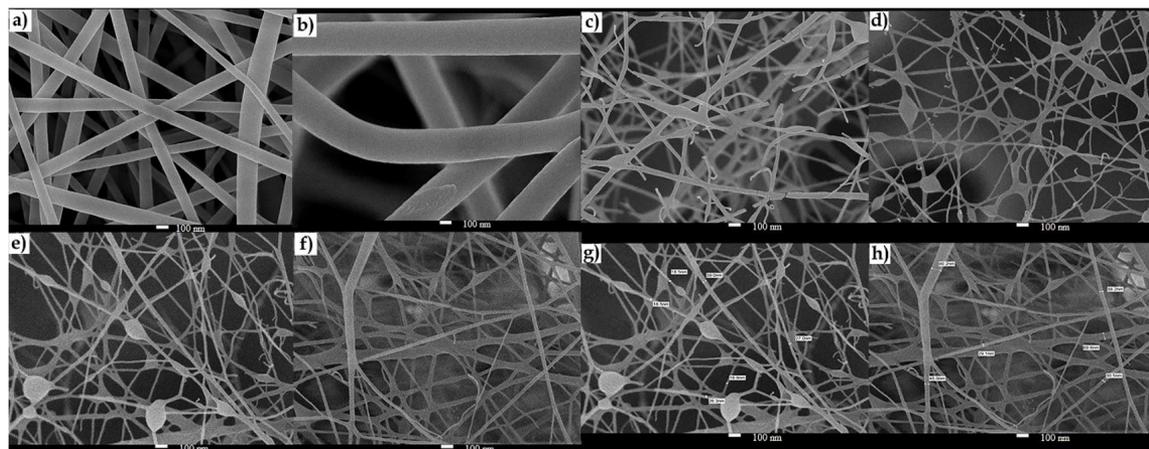


Figure 1. SEM images of (a) sample 1; (b) sample 2; (c) sample 3; (d) sample 4; (e) sample 5; (f) sample 6; (g) sample 7; (h) sample 8.

of the nanofibers increased from 60 to 122 nm when the polymer concentration was increased from 10 to 16%.

3.5 FTIR analysis

FTIR spectrum of 15 and 20% G is presented in Figure 2a. As observed, there is a very slight peak at approximately 3300 cm^{-1} attributed to stretching vibration of amide group (N-H), as well as hydrogen bonding^[24]. Following the signals that represent the digital fingerprint, such as the stretching vibration of: C=O of primary amide, bending and stretching of N-H of secondary amide and finally a bending of de N-H group at 1650 cm^{-1} , 1540 cm^{-1} , and 1250 cm^{-1} , respectively^[38].

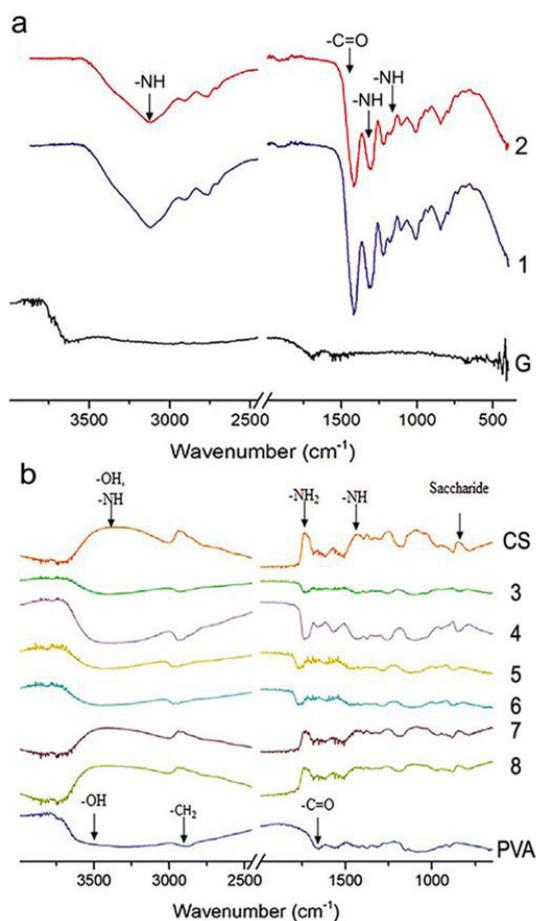


Figure 2. (a) FTIR spectra of G and samples 1 and 2; (b) FTIR spectra of CS-PVA nanofibers and neat CS and PVA.

FTIR spectrum of neat CS and PVA powder, as well as CS-PVA is presented in Figure 2b as well. The FTIR spectra of neat CS showed the characteristic bands of the saccharide structure in the range of 880-1150 cm^{-1} ^[28], as well as the bands for amide I bending at 1660 cm^{-1} and amide II at 1560 cm^{-1} , result of carbonyl stretching by the partial deacetylation of chitin^[29,30]. The signal at 3455 cm^{-1} corresponds to -NH stretching is also present^[31]; however, this band overlaps with the -OH vibrations^[30]. Neat PVA spectra have three predominant signal: The band at 1760 cm^{-1} results of the C=O stretching present on the PVA backbone^[29], the band at 2900 cm^{-1} for the CH_2 asymmetric stretching vibration, and the band about 3500 cm^{-1} attributed to -OH is stretching^[34]. In comparison with neat CS and neat PVA, CS-PVA presents a decrease in the wavenumber of the -OH and -NH signal. This behavior may be credited to the interactions between CS and PVA within the nanofiber system^[30,33]. This interaction is by intermolecular and intramolecular hydrogen bonds through hydrophobic side chain aggregation, as represented by Alhosseini et al.^[39].

The presence of the amino and NAGA signals on the sample is evidence of its biodisponibility, which translates on the theoretical capability of them to carry their antimicrobial and therapeutic effects, by the interaction of the protonated -NH with the microorganism's cell walls^[40] and the stimulation of growth factors by the interaction between the NAGA present in CS and the NAGA receptors in macrophages^[7].

3.6 DSC analysis

DSC data for neat CS, neat PVA, and CS-PVA nanofibers are presented in Table 6. Neat CS presented one endothermic peak at 91.3 $^{\circ}\text{C}$ and PVA an endothermic peak at 181.3 $^{\circ}\text{C}$. However, the samples presented two endothermic peaks, a signal per compound, with a slight displacement of each temperature regarding the neat compounds. This effect denotes an interaction of the compounds, but also a non-miscibility between them. Therefore, it is important to differentiate the concepts of miscibility and interaction of the components. Miscibility is when a single-phase system is formed in a polymer-polymer blend, whereas an interaction is a chemical approach between chemical groups^[37].

3.7 TGA analysis

TGA thermograms of G and CS-PVA electrospun nanofibers are shown in Figure 3. In general terms, the thermal stability of gelatin electrospun fibers was fairly similar to gelatin powder. Beginning with sample 1 (15% of G), it presented a first weight loss at 50 $^{\circ}\text{C}$ to 100 $^{\circ}\text{C}$, which is most likely the result of moisture evaporation. However,

Table 6. Thermal characterization of neat CS and PVA and CS-PVA nanofibers.

Sample	Melting Temperature CS ($^{\circ}\text{C}$)	Melting Temperature PVA ($^{\circ}\text{C}$)	$\Delta\text{H CS}$ (J/g)	$\Delta\text{H PVA}$ (J/g)
Neat	91.3	181.3	350.8	102.9
3	101.0	182.0	70.0	11.5
4	82.9	185.1	67.0	15.0
5	90.3	184.3	44.7	12.0
6	84.1	185.1	83.2	12.8
7	90.0	185.1	56.3	16.3
8	92.4	185.9	60.9	18.5

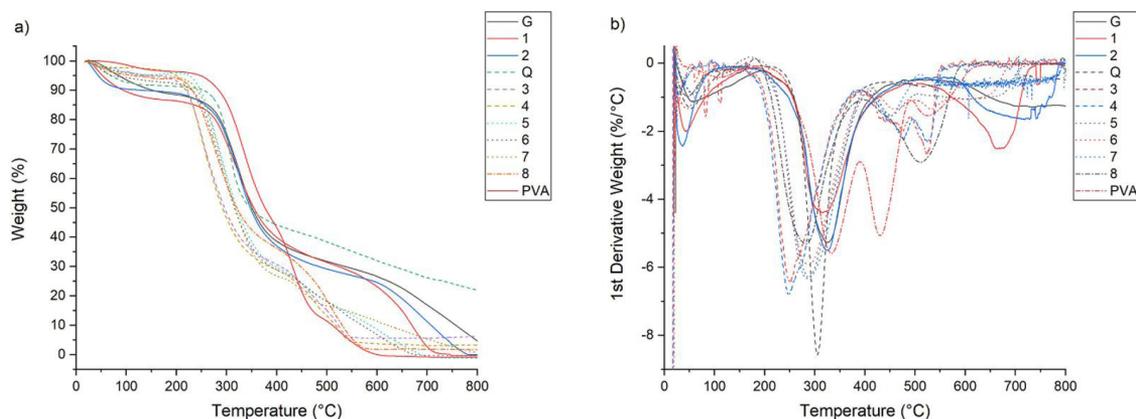


Figure 3. Thermogravimetric curves of (a) TG and (b) DTG of G and CS-PVA nanofibers. G: gelatin, and PVA: polyvinyl alcohol.

around 330 °C the maximum weight loss occurs, which is due to protein degradation^[23]. At high temperatures, above 400 °C corresponds to the thermal decomposition of gelatin networks and molecular arrangements. The changes on thermal stability (maximum temperature) of gelatin samples were corroborated from DTG thermograms (Figure 3b). For CS-PVA samples, the samples showed a release of moisture around 100 °C, although all of these presented a significant weight loss of approximately 80%, from 210 °C to 370 °C. This loss mass is associated with amine unit, paired with $-CH_2OH$ group degradation^[41]. As previously stated on DSC results, there is no apparent miscibility of the polymers, since the material present another twin degradation signal between 310 °C and 420 °C, which is the characteristic of PVA^[42]. It is due to polymer dehydration and by the formation of structures similar to polyacetylene. Moreover, during the thermal decomposition of PVA, it can release CO_2 gas and to form oxides. CS showed the main degradation temperature between 280 to 375 °C, the DTG of Figure 3b indicate that the maximum temperature of CS was 308 °C. Compared with materials, electrospun nanofibers of CS-PVA did not show an improvement on thermal stability. Nevertheless, it is important to emphasize that these results determine a very important part of possible application of these biomaterials. Since it can be observed that the nanofiber can withstand temperatures higher than 100 °C, they can carry out as wound dressings as well as possibly be used as biomedical devices, since these materials generally do not involve a sterilization process with heating above 100 °C.

3.8 Surface hydrophilicity

The contact angle measurement result of water droplets on electrospun CS-PVA nanofiber surfaces are presented in Figure 4. The contact angle of all the nanofibers was found to be $<80^\circ$, which indicates the hydrophilic nature of the material. CS and PVA are hydrophilic materials, were CS hydrophilicity are due to the existence of $-NH$ and $-OH$ groups, as well as PVA hydrophilic nature, is due to its $-OH$ groups^[41,43]. Additionally, an inversely proportional behavior between PVA percentage and contact angle was observed, since at increasing PVA percentage

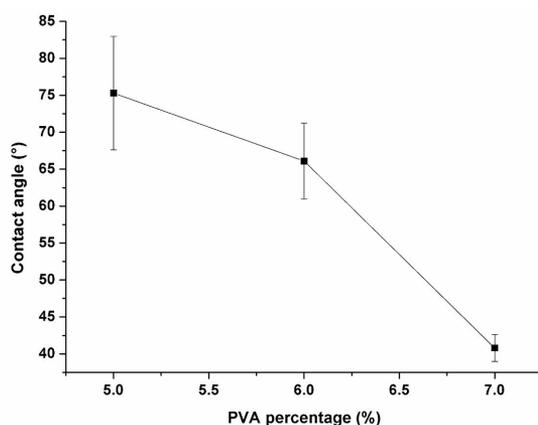


Figure 4. Contact angle for CS-PVA nanofibers.

in the nanofibers, a lower contact angle was obtained. This behavior is because of the hydrophilic nature of PVA explained before. This behavior was previously reported by Agrawal and Pramanik^[44]. The importance of this analysis resides in the fact that, since the nanofiber mats would be for biomedical application. For this end, a hydrophilic material is desired, as previous studies have reported that materials with contact angles between 60 to 80° enhance cell adhesion capability^[39], which is essential for the purpose of this work, that is to say, the application of electrospun CS-PVA nanofibers on diabetic skin ulcers.

3.9 Antimicrobial analysis

Antimicrobial photographs are shown on Figure 5 and Figure 6. With the purpose of screening the possible antimicrobial effect of the nanofibers, bacteria of Gram negative (*S. typhi*) and Gram positive (*S. aureus*) were assayed. It is important to point out that as today, there is no clear activity mechanism for the antimicrobial effect presented by chitosan, and instead, there have been different theories, which will be discussed next. Concerning to the discussion on chitosan and its possible activity mechanism upon *S. typhi*, Verlee et al.^[45], reported two mechanisms which share the characteristic of being developed upon the outer membrane of the bacteria,

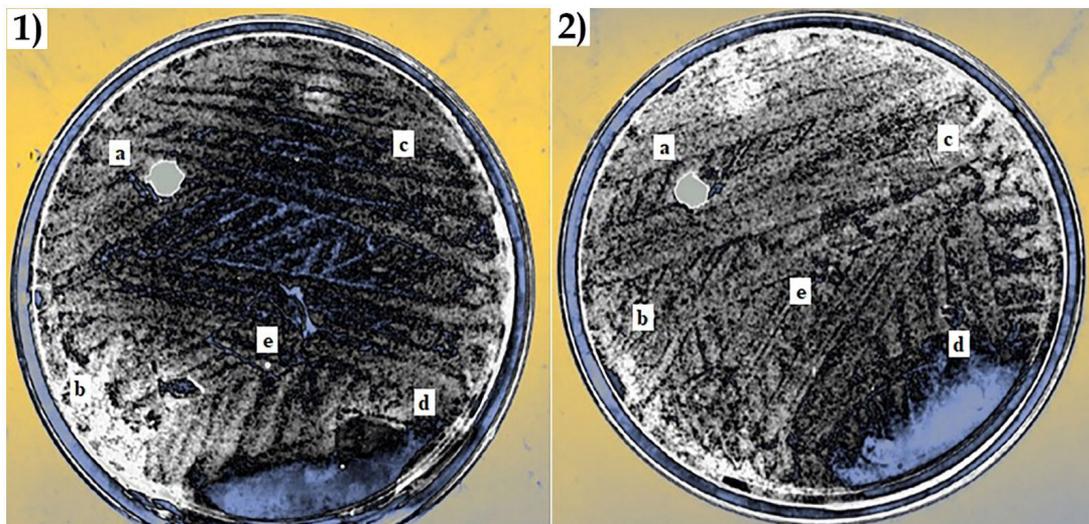


Figure 5. Antimicrobial assay of G and CS-PVA nanofibers against *S. aureus*. 1: (a) control; (b) sample 2; (c) sample 3; (d) sample 4; (e) sample 5. 2: (a) control; (b) sample 1; (c) sample 7; (d) sample 2; (e) sample 8.

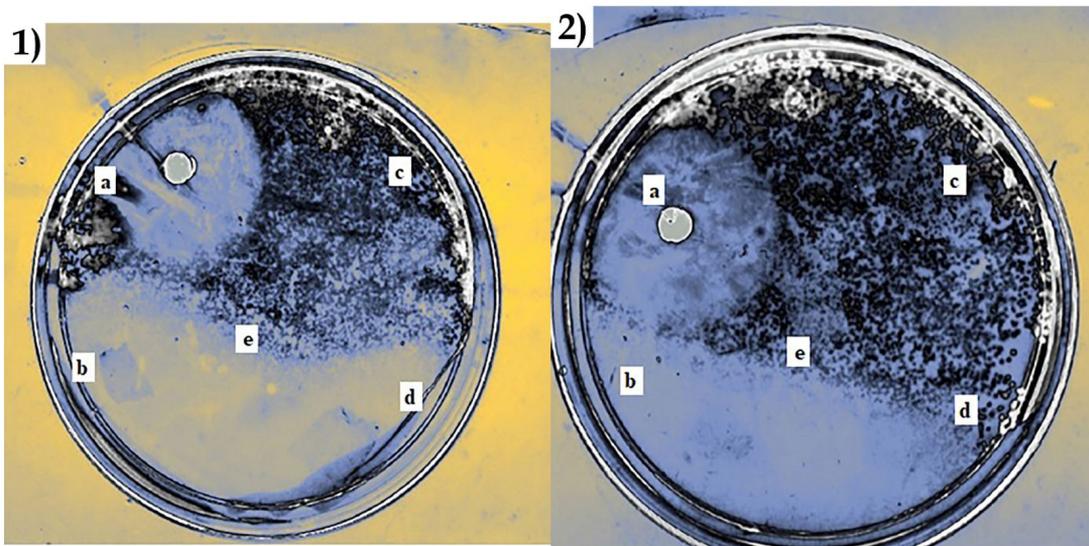


Figure 6. Antimicrobial assay of G and CS-PVA nanofibers against *S. typhi*. 1: (a) control; (b) sample 2; (c) sample 3; (d) sample 4; (e) sample 5. 2: (a) control; (b) sample 1; (c) sample 7; (d) sample 2; (e) sample 8.

chelation of cations (Mg^{2+} , Ca^{2+} , etc)^[46] or electrostatic interaction of chitosan with the lipopolysaccharide present on the outer membrane (principal difference between gram negative and gram positive bacteria). Both effects result on a disruption of the inner membrane^[43], propitiating the intercellular material leaking. All these effects are attributed to the polycationic nature of CS, since in acid media, the amino groups present on CS backbone get protonated. Since membrane configuration of Gram-positive bacteria differs from Gram negative bacteria (as mentioned above), chitosan action mechanism is said to also vary.

As observed on Figure 6, the sample of CS-PVA that presented the most activity was sample 6 and sample 1 for G nanofibers. In the case of these bacteria, such as

S. aureus, the membrane consists, among other things, of teichoic acids embedded into the cell surface. These acids are of the most important for bacterial physiology, since, they are responsible of controlling cationic concentration and enzyme activity, receptor and surfaces binding and protection against environmental stress^[47]. According to Verlee et al.^[45], CS can establish electrostatic interactions with teichoic acids, compromising the bacteria physiology and resulting on antimicrobial activity. The samples that presented the better antimicrobial activity against *S. typhi* were also sample 6 for CS-PVA and sample 1 (15% G). These effects agree with the expressed on the FTIR results, since there are amine group signals either in CS-PVA and G, and both were in acidic media, producing amino protonation.

4. Conclusions

Nanofibers of gelatin and chitosan-polyvinyl alcohol were produced electrospinning process. The electrospun nanofibers produced showed a white color and did not showed chromatic differences between the samples. The gelatin nanofibers showed higher thickness in comparison to CS-PVA nanofibers, it is attributed to complex structure of this biopolymer. The elongation at break of CS-PVA samples is directly related to PVA content. The morphological analysis of gelatin nanofibers and sample 6 showed a homogeneous fibrillar formation. Particular structural and thermal characteristics of the samples were identified by infrared and thermal analysis. The contact angles developed by the samples would be suitable for biomedical applications. The sample that showed the best antimicrobial activity against *S. thyphi*. This sample is capable of integrating with typical small molecules (bioactive compounds) or grow factors to provide a sustained release behavior for an application on chronic patients of diabetes.

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